

# Carbazolothiophene-2-carboxylic acid derivatives as endothelin receptor antagonists

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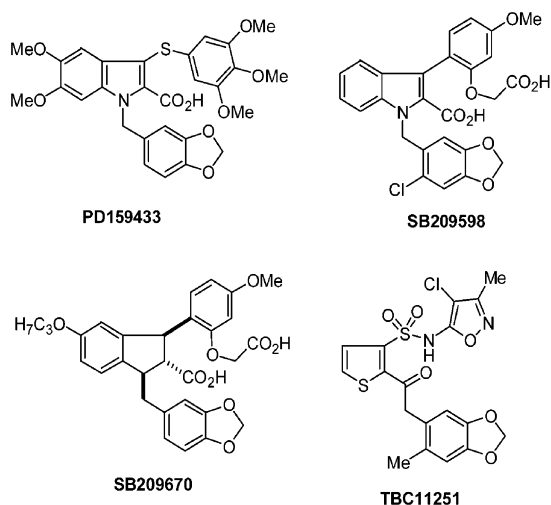
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**Abstract**—The SmI<sub>2</sub>-promoted three-component coupling reaction of thiophene-2-carboxylate, indole-2-carbaldehyde and acetophenone provides an expedient route to a series of tetracyclic carbazolothiophene compounds bearing the indole and thiophene rings. Among these samples, 9-benzyl-4-methyl-4-(4-hydroxyphenyl)-10-oxo-4,10-dihydrocarbazolo[2,3-*b*]thiophene-2-carboxylic acid (**18**) shows the most potent inhibition against the endothelin-1 induced increase of intracellular calcium ion concentration.  
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Human endothelin-1 (ET-1) is a 21 amino acid peptide that exhibits a potent vasoconstrictor activity, conceivably through its selective interaction with specific receptor subtypes (ET<sub>A</sub>, ET<sub>B</sub> and ET<sub>C</sub>).<sup>1</sup> ET-1 contains six highly conserved amino acid residues (His<sup>16</sup>-Trp<sup>21</sup>) at the C-terminus, and this hydrophobic C-terminal hexapeptide alone shows some affinity for ET<sub>A</sub> receptor. Several antagonists including BQ123 [cyclo(L-Leu-D-Val-L-Pro-D-Asp-D-Trp)]<sup>2</sup> are designed on the basis of this peptide structures that incorporate indole moieties. Some non-peptide endothelin antagonists also consist of indole scaffolds such as the indole-2-carboxylic acids PD159433<sup>3</sup> and SB209598<sup>4</sup> (Fig. 1). On the other hand, the molecular modeling indicates that an indan derivative SB209670<sup>5</sup> possesses two phenyl substituents to mimic the amino acid residues of Try-13 and Phe-14 in ET-1. The two carboxylic groups in SB209670 also mimic the Asp-18 residue and the C-terminus of ET-1, which ligate Zn<sup>2+</sup> ion on binding with endothelin receptor.<sup>1</sup> We speculated that a new class of carbazolothiophene derivatives (e.g., **7–22**) bearing appropriate substituents might serve as endothelin receptor antagonists. Indeed, 5-benzyloxy-3-isopropoxy-benzothiophene-2-carboxylic acid<sup>3,6</sup> has been utilized as a lead com-

pound for development of endothelin antagonists. A thiophene-3-sulfonamide TBC11251<sup>7</sup> is also known as an ET<sub>A</sub>-selective antagonist, in which the sulfonamide moiety is considered an isostere of carboxylic acid. We are thus interested in applying our established method of three-component coupling reactions of thiophene-2-carboxylate<sup>8</sup> to synthesize carbazolothiophene-2-



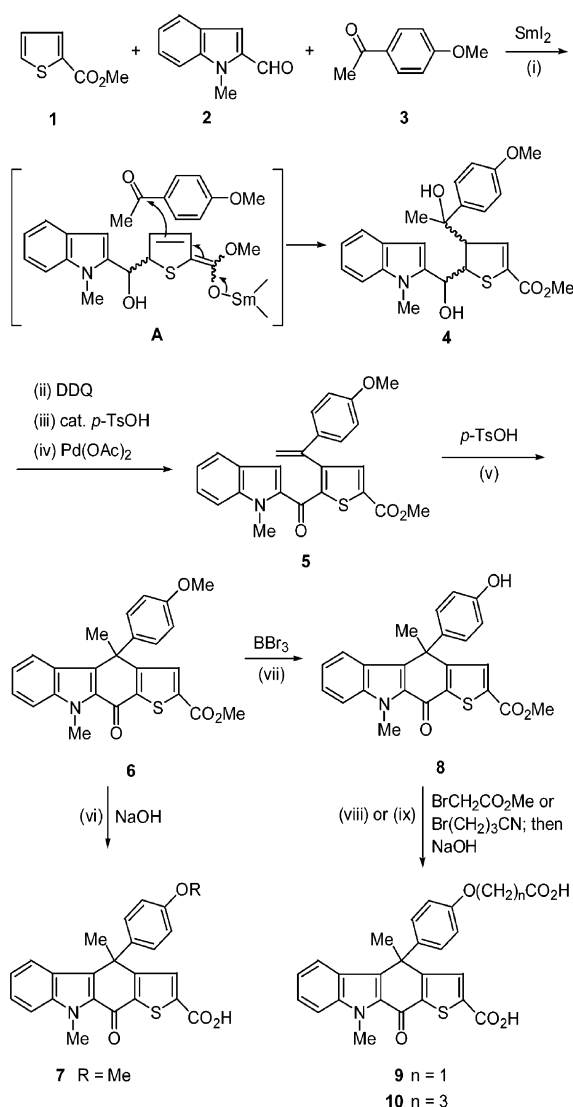
**Figure 1.** Some representative endothelin receptor antagonists constructed by the indole, indan and thiophene scaffolds.

**Keywords:** Coupling reactions; Endothelin; Samarium diiodide; Thiophene; Indole.

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carboxylate derivatives **7–22**, and examined their antagonism against the binding of ET-1 with ET<sub>A</sub> receptor.

By the promotion of samarium diiodide, the three-component coupling reaction of methyl thiophene-2-carboxylate (**1**) with *N*-methylindole-2-carbaldehyde (**2**) and 4-methoxyacetophenone (**3**) occurred smoothly to afford a 77% yield of **4** (Scheme 1).<sup>9,10</sup> This one-pot operation presumably proceeded by an initial coupling of ester **1** with aldehyde **2** to give a dienolate intermediate **A**,<sup>8,11</sup> which was then trapped by ketone **3**. Although diol **4** existed as a mixture of diastereomers, the subsequent oxidation and dehydration would yield a single product. Conversion of **4** to **5** was achieved by a three-step sequence: an oxidation with DDQ to give the

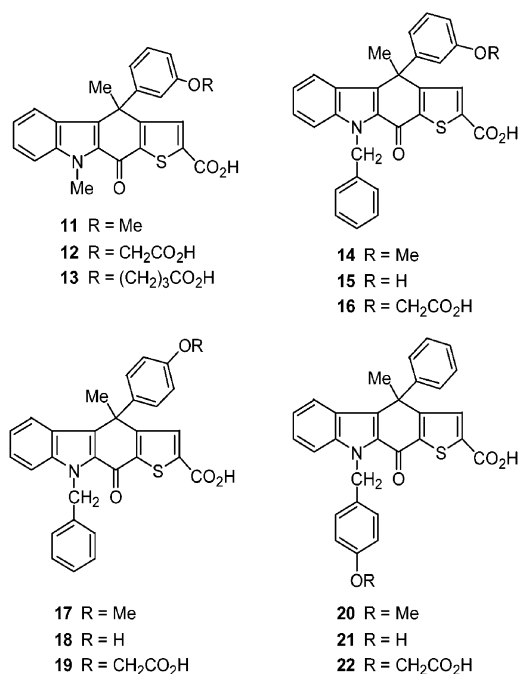


**Scheme 1.** Reagents and conditions: (i) **1** (1 mmol), **2** (1 mmol), SmI<sub>2</sub> (3.6 mmol), THF, HMPA, 0 °C to rt, 1.5 h; then **3** (1.2 mmol), 0 °C to rt, 10 h; 77% yield of **4**. (ii) DDQ, PhH, rt, 4 h; 88%. (iii) cat. *p*-TsOH, PhH, reflux, 10 h; 95%. (iv) Pd(OAc)<sub>2</sub> (5 equiv), K<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>CN, rt, 12 h; 93%. (v) *p*-TsOH, CH<sub>2</sub>Cl<sub>2</sub>, rt, 2 h; 98%. (vi) aq NaOH (0.5%), THF, 0 °C to rt, 2.5 h; 99%. (vii) BBr<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, –78 °C; 96%. (viii) BrCH<sub>2</sub>CO<sub>2</sub>Me, K<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>CN, 80 °C, 15 h; aq NaOH (0.5%), THF, 0 °C to rt, 2.5 h; 95% yield of **9**. (ix) Br(CH<sub>2</sub>)<sub>3</sub>CN, K<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>CN, 80 °C, 48 h; aq NaOH (40%), MeOH, reflux, 1 h; 84% yield of **10**.

corresponding ketone, an acid-catalyzed dehydration to eliminate a water molecule, and an oxidative aromatization by using Pd(OAc)<sub>2</sub> to afford the thiophene product **5**. The subsequent intramolecular Friedel–Crafts alkylation thus furnished the tetracyclic skeleton, giving carbazothienophene-2-carboxylate **6** as a pivotal compound for the synthesis of other derivatives **7–10**. Saponification of **6** afforded acid **7**, whereas demethylation of **6** with BBr<sub>3</sub> gave phenol **8**. Alkylation of phenol **8** with methyl 2-bromoacetate or 4-bromobutanenitrile, followed by hydrolysis in alkaline conditions, gave diacids **9** and **10** in high yields.

A series of carbazothienophenes **11–22** were similarly prepared, initially by the SmI<sub>2</sub>-promoted three-component coupling reactions with appropriate partner substrates. For example, the SmI<sub>2</sub>-promoted coupling product of thiophene ester **1**, indole aldehyde **2** and 3-methoxyacetophenone was further elaborated, according to the procedure similar to that shown in Scheme 1, to give compounds **11–13** bearing MeO, CH<sub>2</sub>CO<sub>2</sub>H or (CH<sub>2</sub>)<sub>3</sub>CO<sub>2</sub>H substituents on the *meta* positions of the phenyl rings as the ‘*meta*-analogues’ of **7**, **9** and **10**. Compounds **14–19** with the (substituted)benzyl groups on the nitrogen atoms were also prepared in high yields by elaboration of the coupling products obtained from ester **1**, 1-benzylindole-2-carbaldehydes and appropriate ketones. Compounds **20–22** are analogues of **17–19** having the substituents switched over. It was noted that the benzyl protons in compounds **14–22** occurred at low fields (ca. δ 6.1) presumably due to the deshielding effect of the adjacent carbonyl group.

The interaction of ET-1 with endothelin receptor is known to trigger an increase of intracellular Ca<sup>2+</sup> concentration ([Ca<sup>2+</sup>]<sub>i</sub>) as the consequence of multistep biological events initiated by G-protein.<sup>12</sup> To evaluate the potency of an endothelin receptor antagonist, one can measure its inhibitory ability against the ET-1



induced  $[Ca^{2+}]_i$  change. According to the reported experimental protocol,<sup>13</sup> Chinese hamster ovary (CHO-K1) cells were transfected with the rat  $ET_A$ -expression plasmid DNA using lipofectin reagent (Life Technologies Inc., USA). The  $ET_A$  overexpression CHO-K1 cells were prior incubated with calcium chelating agent fura-2 applied as its penta(acetoxymethyl) ester,<sup>14</sup> and then treated with ET-1 in  $10^{-7}$  M. The  $[Ca^{2+}]_i$  increase was monitored at 510-nm fluorescence emission by a ratio-metric method using dual excitations at 340 and 380 nm wavelengths.<sup>13</sup> This increment of functional assay was taken as the standard value (100%) to assess the inhibitory potency of compounds **7–22** against the ET-1 binding with receptor. On addition of the test sample **7** ( $10^{-6}$  M) along with ET-1 ( $10^{-7}$  M), only  $30 \pm 5\%$  increment of  $[Ca^{2+}]_i$  was observed (a mean value of three measurements), equivalent to  $\sim 70\%$  inhibition. By comparisons with the known  $ET_A$  antagonists,  $10^{-6}$  M of SB209670 completely inhibited the ET-1 induced  $[Ca^{2+}]_i$ , whereas BQ123 showed  $\sim 60\%$  inhibition under our assay conditions. Accordingly, compounds **9**, **10**, **12**, **13** and **18** showed high inhibition ( $> 75\%$ ) at  $10^{-6}$  M. Compounds **7**, **11**, **15** and **19** showed medium inhibition (50–70%), whereas compounds **14**, **16**, **17** and **20–22** showed low inhibition. Among our examined samples, compound **18** appeared to be the best ET-1 antagonist with  $IC_{50} \sim 10$  nM. In comparison, SB209670 is an even more potent antagonist showing  $\sim 85\%$  inhibition at 10 nM.

In summary, a series of tetracyclic compounds **7–22** bearing the indole and thiophene rings were prepared in an expedient fashion. The functional assay indicated that one of these samples (compound **18**) can serve as a lead compound for future exploration of potent endothelin receptor antagonists. The structure–activity relationship also awaits further investigation.

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- Representative procedure for the  $SmI_2$  promoted three-component coupling reactions: Under an atmosphere of argon, a deep blue  $SmI_2$  solution (0.1 M) was prepared by treatment of Sm (661 mg, 4.4 mmol) with 1,2-diiodoethane (1.01 g, 3.6 mmol) in HMPA<sup>10</sup> (2.8 mL, 16 mmol) and anhydrous THF (32 mL) for 1.5 h at room temperature. To the  $SmI_2$  solution (cooled in an ice bath) were added a THF solution (3 mL) of methyl thiophene-2-carboxylate (142 mg, 1 mmol) and *N*-methylindole-2-carbaldehyde (159 mg, 1 mmol). The reaction mixture was stirred at  $0^\circ C$  for 45 min, and then at room temperature ( $27^\circ C$ ) for 45 min. A THF solution (2 mL) of 4-methoxyacetophenone (180 mg, 1.2 mmol) was added at  $0^\circ C$ , and the mixture was stirred at  $0-27^\circ C$  for additional 10 h. The reaction was quenched by addition of saturated aqueous  $NH_4Cl$  solution (0.1 mL). The mixture was passed through a short silica gel column by rinse with EtOAc/hexane (1:1). The filtrate was concentrated, and chromatographed on a silica gel column by elution with EtOAc/hexane (3:7) to give the desired three-component coupling product **4** (349 mg, 77%) as a mixture of isomers as shown by the  $^1H$  NMR analysis. Compounds **5–22** were fully characterized by spectroscopic methods (IR, MS, HRMS,  $^1H$  and  $^{13}C$  NMR) and elemental analyses. Some pertinent data are listed. **5**:  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.83 (s, 1H), 7.59 (d, 1H,  $J=8.0$  Hz), 7.35 (t, 1H,  $J=8.0$  Hz), 7.24 (d, 1H,  $J=8.0$  Hz), 7.11 (t, 1H,  $J=8.0$  Hz), 6.99 (s, 1H), 6.82 (d, 2H,  $J=8.5$  Hz), 6.59 (d, 2H,  $J=8.5$  Hz), 5.34 (s, 1H), 5.33 (s, 1H), 3.92 (s, 3H), 3.70 (s, 3H), 3.57 (s, 3H). **6**: mp 113–114 $^\circ C$ ;  $^1H$  NMR (400 MHz,  $CDCl_3$ )  $\delta$  7.54 (s, 1H), 7.42 (m, 2H), 7.25 (m, 3H), 7.01 (m, 1H), 6.79 (dd, 2H,  $J=6.8, 2.0$  Hz), 4.28 (s, 3H), 3.86 (s, 3H), 3.74 (s, 3H), 2.09 (s, 3H). **7**: mp  $> 300^\circ C$ . **8**: mp 251–252 $^\circ C$ . **9**: mp 211–212 $^\circ C$ . **10**: mp 261–262 $^\circ C$ . **11**: mp 241–242 $^\circ C$ . **12**: mp 259–260 $^\circ C$ . **13**: mp 271–272 $^\circ C$ . **14**: mp 263–264 $^\circ C$ . **15**: mp 162–163 $^\circ C$ . **16**: mp 149–150 $^\circ C$ . **17**: mp 286–288 $^\circ C$ . **18**: mp 307–308 $^\circ C$ ;  $^1H$  NMR (400 MHz,  $CD_3COCD_3$ )  $\delta$  7.61 (s, 1H), 7.58 (dd, 1H,  $J=8.4, 1.2$  Hz), 7.35–7.18 (m, 9H), 7.02 (dt, 1H,  $J=7.6,$

- 0.8 Hz), 6.78 (td, 2H,  $J=8.8, 3.2$  Hz), 6.16 (d, 1H,  $J=16.0$  Hz), 6.11 (d, 1H,  $J=16.0$  Hz), 2.19 (s, 3H). **19**: mp 244–245 °C. **20**: mp 217–218 °C. **21**: mp 291–292 °C. **22**: mp 150–151 °C.
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